

Doxazosin-Related α_1 -Adrenoceptor Antagonists With Prostate Antitumor Activity

Dario Giardinà,^{*,†} Daniele Martarelli,[‡] Gianni Sagratini,[†] Piero Angeli,[†] Dario Ballinari,[§] Ugo Gulini,[†] Carlo Melchiorre,[⊥] Elena Poggesi,^{||} and Pierluigi Pompei^{*,‡}

[†]Department of Chemical Sciences, University of Camerino, Camerino, Italy, [‡]Department of Experimental Medicine and Public Health, University of Camerino, Camerino, Italy, [§]Department of Oncology, Nerviano Medical Sciences, Nerviano (MI), Italy, ^{||}Pharmaceutical R & D Division, Recordati SpA, Milano, Italy, and [⊥]Department of Pharmaceutical Sciences, University of Bologna, Bologna, Italy

Received December 18, 2008

Doxazosin analogues **1–3** and **1a** were synthesized and investigated at α_1 -adrenoceptors and PC-3, DU-145, and LNCaP human prostate cancer cells. Compound **1** (cyclodoxazosin) was a potent α_{1B} -adrenoceptor antagonist displaying antiproliferative activity higher than that of doxazosin in cancer cells in vitro and in vivo, respectively. Because of its antitumor efficacy at low concentrations, lower apoptotic activity in NHDF vs tumor cells, and antiangiogenic effect, **1** showed a better therapeutic profile relative to doxazosin.

Introduction

Doxazosin is a potent and selective α_1 -adrenoceptor antagonist displaying effective antihypertensive activity.¹ Because of its relaxant activity and increased apoptosis on prostate smooth muscle,^{2,3} it is also used to treat the obstructive and irritative symptoms of BPH^a disease.⁴ Doxazosin exerts, via an apoptotic-induced mechanism, a potent anti-growth effect in vitro against the PC-3, DU-145, and LNCaP human prostate cancer cells, independently of its α_1 -adrenoceptor antagonism or the hormone sensitivity status of cells.^{5,6} The antitumor activity of doxazosin was confirmed in mice bearing PC-3-induced prostate cancer, where it displayed a significant inhibition of tumor growth.⁵ These properties point to a therapeutic significance for the quinazoline-based compounds both in managing human BPH and in the anti-tumor treatment of prostate cancer,⁶ with a possible role in the prevention of tumor development.⁷

In our continuing search for quinazoline-based α_1 -adrenoceptor antagonists, we developed cyclazosin, a racemic compound known to possess high antagonist affinity and selectivity for α_{1A} - and α_{1B} -adrenoceptors relative to the α_{1A} subtype.^{8,9} Here, we report on two novel cyclazosin analogues, 2-[4-(2,3-dihydro-1,4-benzodioxin-2-ylcarbonyl)-*cis*-octahydroquinoxalin-1(2*H*)-yl]-6,7-dimethoxyquinazolin-4-amine (**1**, cyclodoxazosin) and 2-[4-(2,3-dihydro-1,4-benzodioxin-2-ylcarbonyl)-*trans*-octahydroquinoxalin-1(2*H*)-yl]-6,7-dimethoxyquinazolin-4-amine (**2**), incorporating a 2,3-dihydro-1,4-benzodioxine-2-carbonyl moiety instead of the 2-furoyl group, and on the related, previously described,¹⁰ open analogue *N*-{6-[(4-amino-6,7-dimethoxyquinazolin-2-yl) (methyl)amino]hexyl}-*N*-methyl-2,3-dihydro-1,4-benzodioxine-2-carboxamide (**3**), in order to assess the role, if any, of the

decahydroquinoxaline nucleus on the α_1 -adrenoceptor subtypes antagonism and the antitumor activity. In addition, because **1** is more potent than the relative trans isomer **2**, to evaluate the effect of the stereochemistry of the *cis*-octahydroquinoxalin nucleus on the antiproliferative activity (–)-2-[(4*aS*,8*aR*)-4-(2,3-dihydro-1,4-benzodioxin-2-ylcarbonyl)-octahydroquinoxalin-1(2*H*)-yl]-6,7-dimethoxyquinazolin-4-amine (**1a**), a couple of the four stereoisomers of **1**, was included in the study. The stereochemistry of the 1,4-benzodioxine moiety was not investigated because doxazosin was less potent than **1**. The structures of the studied compounds are reported in Figure 1.

Results and Discussion

Chemistry. The synthesis of compounds **1** and **2** was achieved by reaction of 2,3-dihydro-1,4-benzodioxine-2-carbonyl chloride (**4**)¹¹ with *cis*-(**5**)¹² or *trans*-6,7-dimethoxy-2-[octahydroquinoxalin-1(2*H*)-yl]quinazolin-4-amine (**7**), obtained from 2-chloro-6,7-dimethoxyquinazolin-4-amine and *cis*- or *trans*-decahydroquinoxaline,¹³ respectively (Scheme 1). Similarly, **1a** was obtained from **4** and (–)-6,7-dimethoxy-2-[(4*aS*,8*aR*)-octahydroquinoxalin-1(2*H*)-yl]quinazolin-4-amine (**6**).¹⁴ Compound **3** was obtained by reaction of **4** with 6,7-dimethoxy-*N*-methyl-*N*-[6-(methylamino)hexyl]quinazolin-2,4-diamine, as reported.¹⁰ Compounds **1** and **2** are mixtures of four stereoisomers because of the three chiral centers present in their structures. Compound **1** was isolated as mixture of two racemates, in a proportion of 28% and 72%, as shown by the HPLC/DAD analysis, whereas **2** was not analyzed. (see Supporting Information).

α_1 -Adrenoceptor Antagonist Activity and Binding Affinity. Compounds **1–3** and cyclazosin were studied in rat isolated prostatic vas deferens, aorta, and spleen to assess their antagonist activity and selectivity at α_1 -adrenoceptor subtypes.^{15–17} The antagonist potency of cyclazosin at α_{1A} , α_{1B} , and α_{1D} subtypes and of **1** at the α_{1D} subtype was expressed as pA_2 values, according to Arunlakshana and Schild.¹⁸ For **2**, **3**, and for **1** at α_{1A} , and α_{1B} adrenoceptors, as the slope of

*To whom correspondence should be addressed. For D.G.: phone, +39 0737 402265; fax, +39 0737 637345; E-mail, dario.giardina@unicam.it. For P.P.: phone, +39 0737 403314; fax, +39 0737 630618; E-mail, pete.pompei@unicam.it.

^aAbbreviations: BPH, benign prostatic hyperplasia; NHDF, normal human dermal fibroblasts.

the Schild plot was significantly different from unity, the potency was expressed by pK_B values, according to van Rossum.¹⁹ In this latter case, the pK_B value was calculated at the lowest antagonist concentration, giving a significant rightward shift of the agonist concentration–response curve [$\log(\text{concentration ratio} - 1) \geq 0.5$].

The affinity profile of **1–3** and **1a** was also evaluated in radioreceptor binding assays on membranes of Chinese hamster ovary (CHO) cells expressing human α_{1a} , α_{1b} , and α_{1d} adrenoceptor subtypes.²⁰ Binding affinities were expressed as pK_i values derived using the Cheng–Prusoff equation.²¹ The results are reported in Table 1 in comparison with cyclazosin and doxazosin, the reference compounds.

In functional assays, **3** was more potent than **1** and **2** at all α_1 -adrenoceptor subtypes with the exception of **1** at the α_{1B} subtype. Clearly, the insertion of the piperazine ring

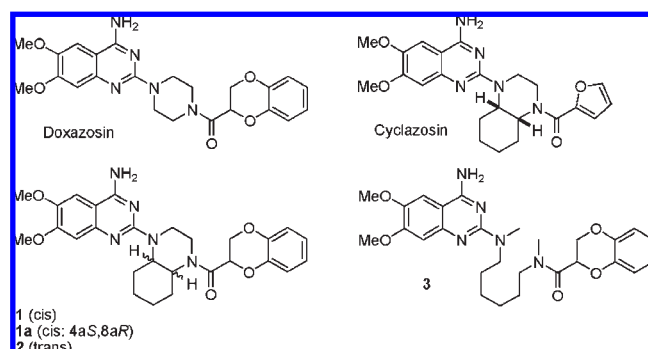
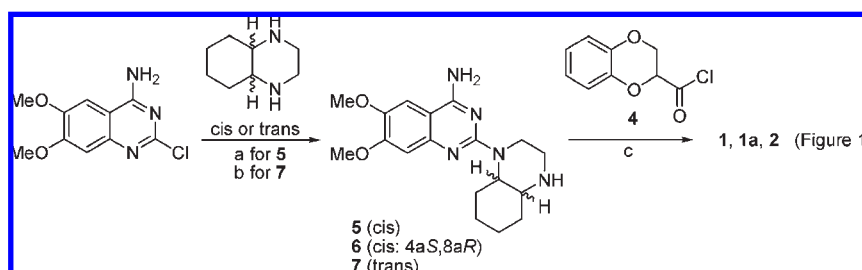


Figure 1. Chemical structures of doxazosin, cyclazosin, **1–3**, and **1a**.

Scheme 1^a



^a Reagents: (a) *i*-AmOH, reflux; (b) *i*-AmOH, triethylamine, DMAP, reflux; (c) dry CH_2Cl_2 .

Table 1. Binding (pK_i) and Functional Affinity (pA_2 or pK_B) Constants and in Vitro Antiproliferative Activity (IC_{50}) of **1–3**, **1a**, Cyclazosin, and Doxazosin at Cloned Human α_1 -Adrenoceptors, Isolated Rat Tissues α_1 -Adrenoceptors, and PC-3, DU-145, and LNCaP Human Prostate Cancer Cells and NHDF Cells^a

no.	pK_i ^a			pA_2 ^b or pK_B ^c			IC_{50} (μM) ^d			
	α_{1a}	α_{1b}	α_{1d}	α_{1A}	α_{1B}	α_{1D}	PC-3	DU-145	LNCaP	NHDF
1	8.55	9.38	9.77	7.21 ± 0.07^c	8.30 ± 0.05^c	7.39 ± 0.06^b	0.85	0.96	0.57	1.46
1a	8.47	8.85	9.31				1.02			
2	7.09	7.65	7.58	5.88 ± 0.18^c	7.14 ± 0.07^c	6.88 ± 0.10^c	$> 10^g$	$> 10^g$	9.75	$> 10^g$
3	8.73	9.03	8.80	8.51 ± 0.07^c	8.35 ± 0.10^c	8.64 ± 0.13^c	8.40	9.25	4.61	9.80
cyclazosin	8.26 ^e	9.49 ^e	9.77 ^e	8.91 ± 0.04^b	9.18 ± 0.03^b	9.21 ± 0.02^b				
doxazosin	9.27 ^f	9.09 ^f	9.09 ^f	$8.69 \pm 0.70^{b,f}$	$9.51 \pm 0.41^{b,f}$	$8.97 \pm 0.23^{b,f}$	38.60	37.44	28.11	43.00

^a Equilibrium dissociation constants (pK_i) were calculated from IC_{50} values using the Cheng–Prusoff equation. The affinity estimates, derived from displacement of [^3H]prazosin binding from α_1 -adrenoceptors, were from two to three experiments performed in triplicate, which agreed within $\pm 20\%$.

^b The α_1 -adrenoceptor blocking activity was assessed by antagonism of (–)-NE-induced contractions of rat vas deferens (α_{1A}) or thoracic aorta (α_{1D}) and by antagonism of (–)-phenylephrine-induced contractions of rat spleen (α_{1B}). pA_2 values are means \pm SEM of three different concentrations, each tested at least four times. ^c pK_B values (\pm SEM) were calculated according to van Rossum. ^d The activity was determined by using a CellTiter-Glo luminescent cell viability assay (Promega). The data, expressed as percentage of control, are reported as IC_{50} values, which represents the concentration (μM) of compounds required for inhibition of 50% of cell growth. Each value is the mean of two independent experiments performed in duplicate, with the standard deviation ranging from 2.4 to 33.5%. ^e Data from ref 12. ^f Data from ref 24. ^g These data indicate that no antiproliferative activity was observed until at least the maximal tested dose (10 μM).

of doxazosin into a *cis*- and *trans*-decahydroquinoxaline nucleus to give **1** and **2**, respectively, is detrimental for the affinity at tissutal α_1 -adrenoceptors. However, the *cis* isomer **1** was more potent than the *trans* **2** at all the subtypes. This is in agreement with the reported higher affinity of cyclazosin relative to its *trans* isomer at the epididymal rat vas deferens α_1 -adrenoceptors.⁸ Notably, unlike **3**, **1** was significantly more potent at the α_{1B} subtype relative to α_{1A} and α_{1D} adrenoceptors (12- and 8-fold, respectively), whereas **2** was more potent at α_{1B} and α_{1D} subtypes with respect to the α_{1A} (18- and 10-fold, respectively).

In the present study, cyclazosin did not display any significant selectivity in functional experiments, in contrast to the results in binding assays.^{9,12} Furthermore, in binding assays, the affinity of **1–3** and cyclazosin at α_1 -adrenoceptor subtypes did not parallel the affinity observed in functional experiments as already found for a number of other prazosin-related compounds.²²

In Vitro Cytotoxic Activity. The antiproliferative effects of **1–3**, **1a**, and doxazosin were evaluated in vitro in PC-3, DU-145, and LNCaP human prostate cancer cells as well as in normal human dermal fibroblasts (NHDF) (Table 1).

Compounds **1** and **3** displayed antiproliferative activity in all tested cancer cells. In particular, **1** exhibited the highest effect, at submicromolar concentrations, whereas **3** was about 10-fold less potent than **1**. The *trans* isomer **2** was 17-fold less potent than **1** at LNCaP cells, and no effect was observed in the other cells up to the maximal tested concentration (10 μM). Compound **1** was 39–49-fold and 8–10-fold more potent on prostate malignant cells than doxazosin and **3**, respectively. Interestingly, **1a** was almost as active as **1**

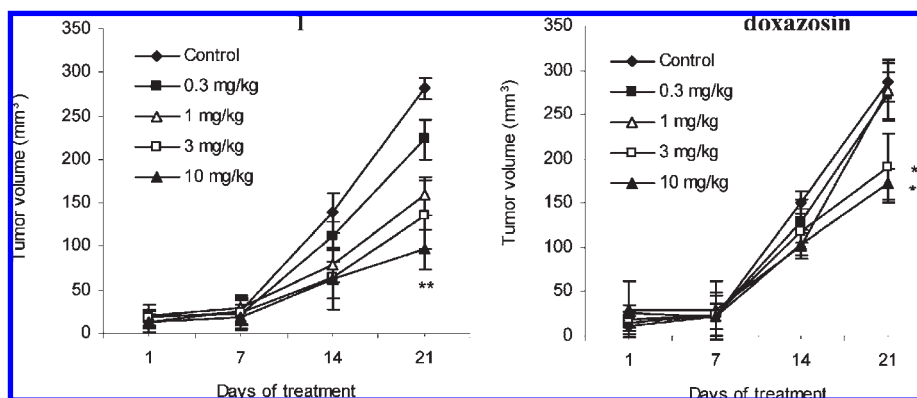


Figure 2. Effect of treatment with **1** and doxazosin on the growth of subcutaneous tumor implanted in male nude mice by human prostate PC-3 cancer cells. Each drug was administered orally and daily, for 21 consecutive days, at specific concentrations to five groups of mice (six animals/group, $n = 6$), under treatment and control. Data represent the mean \pm SEM. Statistical analysis was done by analysis of variance (Anova) and Student t -test; **: significantly different from control, $p < 0.01$.

in PC-3 cells, suggesting that the stereochemistry, at least in these cells, may not have a relevant role in the antiproliferative activity. Concerning **2**, its activity was slightly higher than that of doxazosin in LNCaP cells (3-fold), but not comparable in the others. Like doxazosin, **1–3** displayed an antiproliferative effect at NHDF cells that was close to that shown in prostate cancer lines.

Apoptosis study. Different studies indicated that quinazoline-based α_1 -adrenoceptor antagonists, such as doxazosin, suppress prostate tumor growth via a plurality of mechanisms, including receptor-mediated apoptosis.⁷ Thus, given the structural similarity with doxazosin, the effect of **1–3** on apoptosis was evaluated in PC-3, DU-145, and LNCaP human prostate cancer cells and in NHDF cells through the evaluation of their ability in activation of effector caspases. The level of apoptosis was estimated by measuring in cells the activity of caspase-3 and caspase-7 that are considered the key effector of apoptosis in mammals.²³

Compound **1** was the most effective in caspase induction in cancer cells relative to **2**, **3**, and doxazosin (see Figure 1 of Supporting Information). This finding is in agreement with the higher antiproliferative effect observed for **1** (Table 1). Interestingly, **1**, unlike **2** and **3**, was, at the least, 4–7-fold less effective in inducing caspase activation and apoptosis in NHDF cells ($EC_{50} > 10 \mu\text{M}$) than in tumoral PC-3, DU-145, and LNCaP cells ($EC_{50} = 1.86, 2.60, \text{ and } 1.42 \mu\text{M}$, respectively). On the contrary, the effect of doxazosin at NHDF ($EC_{50} = 40.4 \mu\text{M}$) was similar to that on LNCaP cells but, at the least, 2.5-fold higher than in PC-3 and DU-145 cells ($EC_{50} > 100 \mu\text{M}$).

In Vivo Antitumor Activity. Given its interesting caspase induction ability and antiproliferative activity, **1** was chosen for further in vivo investigation in male nude mice bearing a subcutaneous tumor implanted in the left flank by injection of PC-3 cells. Figure 2 shows the course of tumor inhibition by **1** and doxazosin. A graphical representation of the different efficacy, at each tested concentration at the end of treatment, displayed by **1** and doxazosin, is reported in Figure 2 of Supporting Information.

Compound **1** inhibited the growth of the PC-3 induced tumor in a concentration-dependent fashion, which was significant at the minimal tested dose (21% decrease at 0.3 mg/kg vs control). In contrast, doxazosin gave a significant inhibition only at higher concentrations (3 and 10 mg/kg) (Figure 2 and Figure 2 of Supporting Information).

To confirm the in vitro observed apoptosis induced by **1** and doxazosin, this effect was also investigated in the removed subcutaneous tumor. At the same time, the tumor samples were employed to assess a possible antiangiogenic effect by **1** and doxazosin. The apoptotic effect of **1** and doxazosin was determined by a DNA fragmentation assay in which, through an electrophoretic analysis, a typical DNA ladder pattern was observed on tumor treated with drugs (see Figure 3 of Supporting Information). Again, **1** was more active than doxazosin because, approximately, in tumor treated with **1**, the characteristic pattern was observed at a concentration lower than in doxazosin-treated tumor (0.3 mg/kg vs 3 mg/kg).

The antiangiogenic activity of **1** and doxazosin was determined through immunohistochemical analysis by using the endothelial cell marker rat antimouse CD31 monoclonal antibody. Both drugs affected angiogenesis (see Figure 4 of Supporting Information). At a dose of 3 mg/kg, the drugs reduced blood vessel development. However, tumors treated with **1** were less vascularized than those treated with doxazosin and the effects of **1** on the tumors angiogenesis were also evident at dose of 0.3 and 1 mg/kg.

Conclusions

A few doxazosin analogues (**1–3** and **1a**) were investigated to assess their α_1 -adrenoceptor antagonism and antitumoral effects. Compound **1** displayed a better biological profile than doxazosin and hence a preferential position in potential therapeutic treatment. In particular, **1** was a potent and moderately selective α_{1B} -adrenoceptor antagonist, showing in vitro antiproliferative activity in PC-3, DU-145, and LNCaP human prostate cancer cells, at submicromolar concentrations, and also in vivo antitumor activity in mice PC-3-induced subcutaneous tumor. Although the results do not provide an explanation for the relationship between the α_1 -adrenoceptor antagonism and antitumor activity, compound **1**, bearing a *cis*-octahydroquinoline moiety, might be a useful lead compound for designing new compounds with improved biological profile.

Experimental Section

2-[4-(2,3-Dihydro-1,4-benzodioxin-2-ylcarbonyl)-*cis*-octahydroquinoxalin-1(2*H*)-yl]-6,7-dimethoxyquinazolin-4-amine hydrochloride (1**).** A solution of **4** (1.10 g, 5.54 mmol) in dry CH_2Cl_2

(25 mL) was added dropwise, under nitrogen, to a stirred and cooled (0 °C) solution of **5** (4.20 g, 12.20 mmol) in dry CH₂Cl₂ (75 mL), whereupon the stirring was continued at room temperature for 24 h. Removal of the solvent gave a residue that was purified by silica column chromatography eluting with petroleum ether–ethyl acetate–methanol–14% ammonia (4:4:1:0.1). The obtained free base was transformed into the hydrochloride salt (see Supporting Information for analytical characterization).

(-)-2-[4-(2a*S*,8a*R*)-4-(2,3-dihydro-1,4-benzodioxin-2-ylcarbonyl)-octahydroquinoxalin-1(2*H*)-yl]-6,7-dimethoxyquinazolin-4-amine hydrochloride (**1a**). It was obtained from **4** and **6**¹⁴ following a procedure similar to that described for **1** (see Supporting Information).

2-[4-(2,3-Dihydro-1,4-benzodioxin-2-ylcarbonyl)-*trans*-octahydroquinoxalin-1(2*H*)-yl]-6,7-dimethoxyquinazolin-4-amine hydrochloride (**2**). It was obtained from **4** and **7** following the procedure described for **1** (see Supporting Information).

Acknowledgment. The present work was supported by grants from Fondazione Cassa di Risparmio della Provincia di Macerata (CARIMA), Macerata (Italy), and University of Camerino, Camerino (Italy).

Supporting Information Available: Analytical characterization of **1**, synthetic procedures and analytical characterization of **1a**, **2**, and **7**, elemental analyses, biological methods, in vitro caspase induction and in vivo antitumor efficacy and, apoptotic and antiangiogenic effects of **1** and doxazosin. This material is available free of charge via the Internet at <http://pubs.acs.org>.

References

- Young, R. A.; Brogden, R. N. Doxazosin: A review of its pharmacodynamic and pharmacokinetic properties and therapeutic efficacy in mild or moderate hypertension. *Drugs* **1988**, *35*, 525–541.
- Chapple, C. R. Pharmacotherapy for benign prostatic hyperplasia—the potential for α_1 -adrenoceptor subtype-specific blockade. *Br. J. Urol.* **1998**, *81* (Suppl. 1), 34–47.
- Chon, J.; Borkowski, A.; Partin, A. W.; Isaacs, J. T.; Jacobs, S. C.; Kyprianou, N. α_1 -Adrenoceptor antagonists terazosin and doxazosin induce prostate apoptosis without affecting cell proliferation in patients with benign prostatic hyperplasia. *J. Urol.* **1999**, *161*, 2002–2008.
- Djavan, B.; Marberger, M. A meta-analysis on the efficacy and tolerability of alpha-1-adrenoceptor antagonists in patients with lower urinary tract symptoms suggestive of benign prostatic obstruction. *Eur. Urol.* **1999**, *36*, 1–13.
- Kyprianou, N.; Benning, C. M. Suppression of human prostate cancer cell growth by α_1 -adrenoceptor antagonists doxazosin and terazosin via induction of apoptosis. *Cancer Res.* **2000**, *60*, 4550–4555.
- Benning, C. M.; Kyprianou, N. Quinazoline-derived α_1 -adrenoceptor antagonists induce prostate cancer cell apoptosis via an α_1 -adrenoceptor-independent action. *Cancer Res.* **2002**, *62*, 597–602.
- Garrison, J. B.; Shaw, Y.-J.; Chen, C.-S.; Kyprianou, N. Novel quinazoline-based compounds impair prostate tumorigenesis by targeting tumor vascularity. *Cancer Res.* **2007**, *67*, 11344–11352.
- Giardinà, D.; Gulini, U.; Massi, M.; Piloni, M. G.; Pompei, P.; Rafaiiani, G.; Melchiorre, C. Structure–activity relationships in prazosin-related compounds. 2. Role of the piperazine ring on α -blocking activity. *J. Med. Chem.* **1993**, *36*, 690–698.
- Giardinà, D.; Crucianelli, M.; Melchiorre, C.; Taddei, C.; Testa, R. Receptor binding profile of cyclazosin, a new α_{1B} -adrenoceptor antagonist. *Eur. J. Pharmacol.* **1995**, *287*, 13–16.
- Giardinà, D.; Crucianelli, M.; Marucci, G.; Melchiorre, C.; Polidori, C.; Pompei, P.; Massi, M. Pharmacological evaluation of prazosin- and doxazosin-related compounds with modified piperazine ring. *Arzneim.-Forsch. Drug Res.* **1996**, *46*, 1054–1059.
- Koo, J.; Avakian, S.; Martin, G. J. Derivatives of 1,4-benzodioxan. I. 1,4-Benzodioxan-2-carboxamides. *J. Am. Chem. Soc.* **1955**, *77*, 5373–5374.
- Giardinà, D.; Polimanti, O.; Sagratini, G.; Angeli, P.; Gulini, U.; Marucci, G.; Melchiorre, C.; Poggesi, E.; Leonardi, A. Searching for cyclazosin analogues as α_{1B} -adrenoceptor antagonists. *Il Farmaco* **2003**, *58*, 477–487.
- Brill, E.; Schultz, H. P. Quinoxaline studies. XII. Stereodirective syntheses of *cis*- and *trans*-decahydroquinoxalines and *cis*- and *trans*-decahydroquinoxalones-2. *J. Org. Chem.* **1964**, *29*, 211–220.
- Sagratini, G.; Angeli, P.; Buccioni, M.; Gulini, U.; Marucci, G.; Melchiorre, C.; Leonardi, A.; Poggesi, E.; Giardinà, D. Synthesis and α_1 -adrenoceptor antagonist activity of derivatives of the furan portion of (+)-cyclazosin. *Bioorg. Med. Chem.* **2007**, *15*, 2334–2345.
- Eltze, M.; Boer, R.; Sanders, K. H.; Kolassa, N. Vasodilatation elicited by 5-HT_{1A} receptor agonists in constant-pressure-perfused rat kidney is mediated by blockade of α_{1A} -adrenoceptors. *Eur. J. Pharmacol.* **1991**, *202*, 33–44.
- Ko, F. N.; Guh, J. H.; Yu, S. M.; Hou, Y. C.; Wu, Y. C.; Teng, C. M. Discretamine, a selective α_{1D} -adrenoceptor antagonist, isolated from *Fissistigma glaucescens*. *Br. J. Pharmacol.* **1994**, *112*, 1174–1180.
- Buckner, S. A.; Oheim, K. W.; Morse, P. A.; Knepper, S. M.; Hancock, A. A. Alpha 1-adrenoceptor-induced contractility in rat aorta is mediated by the alpha 1D subtype. *Eur. J. Pharmacol.* **1996**, *297*, 241–248.
- Arunlakshana, D.; Schild, H. O. Some quantitative uses of drug antagonists. *Br. J. Pharmacol.* **1959**, *14*, 48–58.
- van Rossum, J. M. Cumulative dose–response curves. II. Technique for the making of dose–response curves in isolated organs and the evaluation of drugs parameters. *Arch. Int. Pharmacodyn. Ther.* **1963**, *143*, 299–330.
- Testa, R.; Taddei, C.; Poggesi, E.; Destefani, C.; Cotecchia, S.; Hieble, J. P.; Sulpizio, A. C.; Naselsky, D.; Bergsma, D.; Ellis, C.; Swift, A.; Ganguly, S.; Ruffolo, R. R., Jr.; Leonardi, A. A novel prostate selective α_1 -adrenoceptor antagonist. *Pharmacol. Commun.* **1995**, *6*, 79–86.
- Cheng, Y.-C.; Prusoff, W. H. Relationship between the inhibition constant (K_i) and the concentration of inhibitor which causes 50% inhibition (IC_{50}) of an enzyme reaction. *Biochem. Pharmacol.* **1973**, *22*, 3099–3108.
- Antonello, A.; Hrelia, P.; Leonardi, A.; Marucci, G.; Rosini, M.; Tarozzi, A.; Tumiatti, V.; Melchiorre, C. Design, synthesis, and biological evaluation of prazosin-related derivatives as multipotent compounds. *J. Med. Chem.* **2005**, *48*, 28–31.
- Hanahan, D.; Weinberg, R. A. The hallmarks of cancer. *Cell* **2000**, *100*, 57–70.
- Hancock, A. A.; Buckner, S. A.; Brune, M. E.; Katwala, S.; Milicic, I.; Ireland, L. M.; Morse, P. A.; Knepper, S. M.; Meyer, M. D.; Chapple, C. R.; Chess-Williams, R.; Noble, A. J.; Williams, M.; Kerwin, J. F., Jr. Pharmacological characterization of A-131701, a novel α_1 -adrenoceptor antagonist selective for α_{1A} and α_{1D} compared to α_{1B} -adrenoceptors. *Drug Development Research* **1998**, *44*, 140–162.